MODELS OF FOLATE COFACTORS - 22.¹ LEWIS ACID CATALYZED CYCLIZATION OF CARBON-FRAGMENT TRANSFER PRODUCTS OF FOLATE COFACTOR MODELS. SYNTHESIS OF ENANTIO-MERICALLY PURE TETRACYCLIC (ABCE) RING SYSTEM OF ASPIDOSPERMA ALKALOIDS.

R.H. Huizenga² and U.K. Pandit^{*} Organic Chemistry Laboratory, University of Amsterdam, Nieuwe Achtergracht 129, 1018 WS Amsterdam, The Netherlands

(Received in UK 29 January 1991)

Abstract: Michael adducts of tryptamine, N(1)-methyltryptamine and tryptophane with acrylic esters react with substituted imidazolidines (N(5),N(10)-methylenetetrahydrofolate models) to give enaminones which cyclize under influence of Lewis acid, to give mixtures of tetrahydro-1*H*-pyrido[3,4-b]indoles and pyrrolo[2,3-d]carbazoles. The influence of the nature of Lewis acid on product formation is discussed. The synthesis of an enantiomerically pure pyrrolo[2,3-d]carbazole system is reported.

Work in our laboratory has demonstrated that suitable folate cofactor models can be employed as pivotal reagents in the synthetic approach to a variety of natural products and their analogues.³ In our studies directed to the application of substitued N(5),N(10)-methylenetetrahydrofolate models in alkaloid synthesis, we have reported the preparation of heterocyclic systems representing the functionalized Aspidosperma skeleton.^{1,4} In this study the folate cofactor models have been so chosen that the carbon-fragment transferred, contains the carbon atoms 16, 17, 20, and 21 of the final Aspidosperma skeleton as well as the alkoxycarbonyl group at C(16), which is present in several, pharmacological active, members of the Aspidosperma alkaloid family.⁵ We emphasize our results on the Lewis acid induced cyclization of the enaminone intermediates derived from the reaction of tryptamine derivatives **3a,b** and tryptophanate ester **27**, with folate cofactor model **4**. In this cyclization reaction, which is analogous to that utilized in several synthetic strategies towards the Aspidosperma skeleton, the C(7)-C(21), and the C(2)-C(16) bonds are formed, presumably in a stepwise manner, to generate rings C and E of the alkaloid system.^{6,7}

The starting substances 3a,b were conveniently obtained by the Michael addition of tryptamines 1a,b to acrylic esters (2a,b) (Scheme 1). The carbon-fragment transfer from the folate model, employed in its ring-opened form 5, to 3a,b, furnished the crucial enaminone intermediates 6a,b. These were subjected to cyclization reactions under influence of BF₃·OEt₂ or TiCl₄, whereupon tetrahydro- β -carbolines 7a,b and octahydro-1H-pyrrolo-[2,3-d]carbazoles 8a,b were formed as the salient reaction products.⁸ The reaction mixtures could be monitored and analyzed by HPLC and the results are presented in Table 1.

| Table 1 |
|--|
| Lewis acid catalyzed cyclization of 6a,b |

| compound | Lewis acid | pyridoindole | pyrrolocarbazole |
|----------|-------------------|--------------|------------------|
| 6a | BF3·OEt2 | 7a 45 % | 8a 19 % |
| 6 b | BF3-OEt2 | 7b 49 % | - |
| 6a | TiCl ₄ | - | 8a 26 % |
| 6b | TiCl4 | - | 8b 55 % |



Inspection of Table 1 reveals that the BF₃-OEt₂ catalyzed cyclization results predominantly in the pyridoindole system, whereas the TiCl₄ mediated reaction leads exclusively to the tetracyclic pyrrolocarbazole product. The mechanism of formation of the two products can be explained by considering, in particular, the coordination of the Lewis acid with the two carbonyl groups of the β -keto ester moiety in the enaminone system 6 (a,b). The resulting iminium intermediate 9 (a,b) (Scheme 2), undergoes a cyclization step involving the indole nucleus as a nucleophile, whereupon intermediate 10 (a,b) is formed. This spiroindolenine system can undergo two different fates depending upon the nature of the Lewis acid. According to pathway [a], the intermediate 10 (a,b) undergoes a typical Wagner-Meerwein rearrangement, to result in the β -carboline cation 11 (a,b) which deprotonates to form 7 (a,b). In an alternate pathway [b], 10 (a,b) can isomerize to 12 (a,b), which subsequently cyclizes to tetracyclic pyrrolocarbazole 8 (a,b), via intermediate 14 (a,b).

In the BF₃·OEt₂ mediated reaction, processes [a] and [b] compete with each other, the product distribution depending upon the relative stabilities of systems 10 and 12. It should be noted that in case of 12, complexation with BF₃·OEt₂ will stereoelectronically favour a situation in which the two BF₃ coordinated sites are remote from each other (E-configuration; e.g. 13). However, in 13, the transition state of the cyclization step will encounter serious steric repulsion between the indole nucleus and the BF₃-complexed ester function. This factor will adversely affect the formation of the tetracyclic heterocyclic system 8 (a,b). This is in line with the observation that, whereas 6a yields 8a as the minor product, the introduction of the methyl substituent in the indole moiety (6b) presumably suppresses intermediates 12b = 13b, that is pathway [b], and thereby the formation of 8b.



The exclusive formation of tetracyclic products 8a and 8b in the TiCl₄ induced cyclization reactions is attributed to the ability of Ti(IV) to coordinate simultaneously with more than one electro-negative group. The same Ti(IV) cation will coordinate with both the oxygen of the enolate and the carbonyl group of the ester. This results in the formation of intermediate 14 (a,b), in which there is minimum steric hindrance to the cyclization reaction, leading to 8 (a,b). The importance of steric interactions in this cyclization step is emphasized by the TiCl₄ induced cyclization reaction of enaminone 18 (Scheme 3).



Open form of the folate model i.e. 17 was formed upon addition of the imidazolidinium salt 15 to the dianion of β -keto ester (16). The carbon-fragment transfer reaction from 17 to tryptamine derivative 3b furnished enaminone 18, that bears an ethyl substituent on the α -carbon of the β -keto ester. Treatment of enaminone 18 with TiCl4 led only to the isolation of the β -carboline 19. This is clearly due to additional steric hindrance between the ethyl group and the indole nucleus in the spiroindolenine intermediate 20; the attack of the enolate on the iminium ion being prevented, the intermediate reverts to the one corresponding to 10 (Scheme 1), which after a Wagner-Meerwein rearrangement leads to 19.

The stereochemistry of **8a** and **8b** was established by NOE experiments. Irradiation of the signal of $C(2)H_{\beta}$ of **8a** resulted in an enhancement of the signal ascribed to $C(6)H_{\beta}$, and irradiation of the signal of C(21)H resulted in an enhancement of a doublet at 6.7 ppm ascribed to C(9)H. In compound **8b** a nuclear Overhauser effect was observed for the protons at C(5) and C(21) upon irradiation of the signal of C(9)H. Irradiation of the signal of $C(2)H_{\beta}$ gave an enhancement of the signals of $C(6)H_{\beta}$ and NMe.

The tetracyclic ring systems have the same stereochemistry as the Aspidosperma alkaloids, viz. that the B/C and C/E ring junction possess the *cis* configuration. The stereochemistry of the reaction is determined during the formation of the spiro intermediate (e.g. 22 and 25 in Scheme 4). Two conformations of the iminium enolate (21 and 24) can be visualized, as is shown in Scheme 4. In intermediate 21 the indole nucleus lies above the iminium double bond. Formation of the spiro-intermediate 22, followed by the second cyclization furnishes the indolenine 23 with the *trans* C/E ring junction. In intermediate 24 the iminium double bond of the indole nucleus here the indole nucleus have a rans configuration. Nucleophilic attack of the enamine double bond of the indole followed by the second cyclization step results in compound 26, with the correct *cis* C/E geometry. It is evident that because of steric repulsion, the intermediate 24, in which the enolate and indolylethyl substituent (of the iminium double bond) have a *trans*-configuration, is favoured over intermediate 21. The selective formation of



the less strained tetracycles with the cis C/E ring junction can also be explained by assuming the reversability of the spiro intermediate formation.⁹ The product formation can then be regarded to be thermodynamically controlled.

To see whether the asymmetric centre in L-(-)-tryptophane is capable of inducing stereospecificity in the TiCl4 induced cyclization reaction, tryptamine was substituted by L-(-)-tryptophane methyl ester 27 in an analogous reaction scheme (Scheme 5). The Michael addition of 27 to methyl acrylate furnished compound 28. Transfer of the carbon fragment of ring-opened model 29 to this compound resulted in enaminone 30. This enaminone was optically active $([\alpha]^{20}_{D} = -121^{\circ} (c = 0.0102, CHCl_3))$. The cyclization reaction of 30 with 2 eq TiCl4 provided the tetracyclic ring system 31 $([\alpha]^{20}_{D} = -50.3^{\circ} (c = 0.0167, CHCl_3))$. Only one product was isolated. Assuming that there had been no epimerization at C(5), it has the absolute stereochemistry shown in 31; which is identical with the natural vindoline template. The ¹H-NMR spectrum of 31 was assigned unambiguously using the 2D-COSY technique. Furthermore, with NOE experiments the stereochemistry of the molecule was established. Irradiation of the signal of C(2)Hg gave an enhancement of the signals of C(6)Hg (dd, 2.08 ppm), the NH and C(16)H (m, 4.90 ppm). No interaction with C(21)H could be observed, which implies that C(21)H possesses the α orientation. The β orientation of the methoxycarbonyl group at C(5)H can be concluded from the nuclear Overhauser effect observed between C(5)H and C(21)H_α.

The enantiomeric purity of 31 was established by ¹H-NMR. Addition of the chiral shift reagent $Eu(hfc)_3$ to a solution of 31 did not result in splitting of signals in the ¹H-NMR spectrum. A complex was formed between $Eu(hfc)_3$ and 31, as was attested by the shift of the signals in the spectrum of 31.



Scheme 5

Experimental

Chromatographic separations were carried out by means of flash chromatography on freshly filled suica gel (230-400 mesh) columns, following literature procedure.¹⁰ All m.ps are uncorrected. Infrared spectra were recorded on a Perkin Elmer 257 or 298 spectrometer. The absorbtions are given in cm⁻¹. ¹H-NMR measurements were performed on Varian A-60, HA-100 or XL-100 instruments or on Brucker WM-250 or AC-200 instruments. ¹³C-NMR spectra were recorded on the Brucker WM-250 or AC-200 instruments. The chemical shifts are given in ppm downfield from tetramethylsulane. Unless stated otherwise IR and NMR spectra are taken in CHCl₃ and CDCl₃, respectively. Exact mass measurements were carried out using a Varian MAT 711 or a VG Micromass ZAB-2HF. The IUPAC nomenclature is used in naming the compounds. In the text and in the description of the NMR spectra a numbering method related to the Aspidosperma alkaloids is used (See note 7).

Methyl 3-(2-(3-indolyl)ethyl)aminopropionate (2a R₁ = Me).

A solution of methyl acrylate (25 mmol, 2.1 g) in 20 ml abs. ethanol was added dropwise to a solution of 25 mmol (4.0 g) tryptamine in 50 ml of ethanol. The mixture was stirred for 2 h at room temperature and heated to 75 °C for 0.5 h. The mixture was concentrated under vacuum and purified by flash chromatography (ethyl acetate \rightarrow ethyl acetate/ethanol 1:1). 5.98 g (93 %) of the product was isolated as a dark oil. ¹H-NMR (250 MHz): 1.78 (br.s, 1H, CH₂NHCH₂), 2.50 (t, 2H, J = 6.6 Hz, CH₂CO₂), 2.96 (br.s, 4H, indole-CH₂CH₂NH), 3.60 (s, 3H, OCH₃), 6.98 (d, 1H, J = 2.3 Hz, C(2)H indole), 7.07-7.21 (m, 2H, C(5)H and C(6)H indole), 7.31 (d, 1H, J = 7.7 Hz, C(7)H indole), 7.61 (d, 1H, J = 7.7 Hz, C(4)H indole), 8.43 (br.s, 1H, NH indole). IR: 3480 (sh.s), 3300 (br.w), 1730 (s).

Ethyl 3-(2-(3-indolyl)ethyl)aminopropionate (2b R₁ = Et).

Ethyl acrylate (15 mmol, 1.59 g) was added dropwise to a solution of 15 mmol tryptamine in 25 ml of abs. ethanol at 0 °C. After stirring for 24 h the reaction mixture was concentrated under vacuum and purified by flash chromatography (ethyl acetate \rightarrow ethyl acetate-ethanol 1:1). The product was obtained as a red brown gum 83 % yield (3.25 g). Crystallization from ethyl acetate furnished yellow crystals (mpt. 71-72.5 °C). ¹H-NMR (250 MHz): 1.13 (t, 3H, J = 7.2 Hz, OCH₂CH₃), 2.00 (br.s, 1H, CH₂NHCH₂), 2.28-2.40 (m, 2H, CH₂CO₂), 2.80-3.05 (m, 6H, CH₂CH₂NCH₂CH₂CO₂), 4.05 (q, 2H, J = 7.2 Hz, OCH₂CH₃), 6.90-7.70 (m, 5H, indole), 8.80 (br.s, 1H, NH indole). ¹³C-NMR (50 MHz, APT): 13.9 (OCH₂CH₃), 25.4 (indole-CH₂CH₂), 34.4 (CH₂CO₂), 44.7 and 49.5 (CH₂NHCH₂), 60.2 (OCH₂CH₃), 111.0 (C(7) indole), 113.0 (C(3) indole), 118.5 and 118.8 (C(4) and C(6) indole), 121.6 and 122.0 (C(5) and C(2) indole), 127.1 (C(3a) indole), 136.2 (C(7a) indole), 172.6 (CO₂Et). IR: 3480 (sh.s), 3300 (br.w), 1725 (s).

Ethyl 3-(2-(3-(N-methyl))indolyl)ethyl)amunopropionate (3b R₁ = Et).

A solution of 6.0 mmol (603 mg) ethyl acrylate in ethanol was added dropwise to a solution of 6.0 mmol N(1)-methyltryptamine in 10 ml abs. ethanol at room temperature under a nitrogen atmosphere. The solution was heated to reflux and after 3 h concentrated under vacuum and purified by flash chromatography (ethyl acetate \rightarrow ethyl acetate/ethanol 9:1). 1.413 g (85 %) of the product was isolated as a dark oil. ¹H-NMR (200 MHz): 1.26 (t, 3H, J = 7.1 Hz, OCH₂CH₃), 2.49-2.52 (m, 2H, CH₂CO₂), 2.85-3.05 (m, 6H, CH₂CH₂NHCH₂), 3.76 (s, 3H, OCH₃), 4.12 (s, 3H, NCH₃), 6.89 (s, 1H, C(2)H indole), 7.10-7.35 (m, 3H, C(5)H, C(6)H and C(7)H indole), 7.63 (d, 1H, J = 7.6 Hz, C(4)H indole). ¹³C-NMR (50 MHz, APT): 14.0 (OCH₂CH₃), 25.4 (indole-CH₂CH₂NH), 32.3 (NCH₃), 34.6 (CH₂CO₂), 44.8 and 49.8 (CH₂NHCH₂), 61.1 (OCH₂CH₃), 109.0 (C(7) indole), 112.2 (C(3) indole), 118.5 and 118 7 (C(4) and C(6) indole), 121.4 (C(5) indole), 126.5 (C(2) indole), 127.7 (C(3a) indole), 136.9 (C(7a) indole), 172.4 (CO₂Et). IR: 1730 (s). MS (FD 10 mA): 274.

Methyl 3-(2-(3-(N-methyl))indolyl)ethyl)aminopropionate (3b R₁ = Me).

A solution of 2.61 g (28.7 mmol) methyl acrylate in 20 ml of ethanol was slowly added to a solution of 5.0 g (28.7 mmol) N(1)methyltryptamine. The mixture was stirred for another 2 h at room temperature and heated for 0.5 h to 60 °C. After concentration of the mixture under vacuum it was purified by flash chromatography (ethyl acetate \rightarrow ethyl acetate/ethanol 7:3) and 5.92 g (79 %) product was obtained as a brown oil. ¹H-NMR (250 MHz): 2.06 (s, 1H, NH), 2.51 (t, 2H, J = 6.7 Hz, CH₂CO₂), 2.92 (t, 2H, J = 6.7 Hz, NHCH₂), 2.95 (s, 4H, indole-CH₂CH₂NHCH₂), 3.62 (s, 3H, OCH₃), 3.72 (s, 3H, NCH₃), 6.88 (s, 1H, C(2)H indole), 7.07-7.33 (m, 3H, C(5)H, C(6)H and C(7)H indole), 7.61 (d, 1H, J = 7.8 Hz, C(4)H indole). ¹³C-NMR (50 MHz, APT): 25.4 (indole-CH₂CH₂NH), 32.4 (NCH₃), 34.4 (CH₂CO₂), 44.8 and 49.8 (CH₂NHCH₂), 51.4 (OCH₃), 109.1 (C(7) indole), 112.1 (C(3) indole), 118.6 and 118.8 (C(4) and C(6) indole), 121.4 (C(5) indole), 126.6 (C(2) indole), 127.7 (C(3a) indole), 136.9 (C(7a) indole), 173.0 (CO₂Me), IR: 1725 (s).

Ethyl 5-[ethoxycarbonylethyl-(3-indolyl)ethyl]amino-3-oxo-4-pentenoate (6a R1=Et).

A mixture of tryptamine derivative 3a (2600 mg, 10 mmol) and folate model 5 (750 mg, 2 mmol) were refluxed in 16.5 ml of acetonitrile/acetic acid (10:1) under a nitrogen atmosphere for 6.5 h. The reaction mixture was concentrated under vacuum and the residue was dissolved in ethyl acetate. After addition of some silica gel the solvent was removed under reduced pressure and the powder was brought on top of a silica gel column for flash chromatography (eluent ethyl acetate/petroleum ether 60-80 1:1 \rightarrow ethyl acetate) The product was isolated as a yellow oil in 63 % yield (496 mg). ¹H-NMR (250 MHz) broad signals due to hindered rotation: 1.21-1.27 (m, 6H, 2 x O₂CH₂CH₃), 2.02-2.03 (m, 2H, NCH₂CH₂O₂), 2.40-2.51 (m, 2H, indole-CH₂), 2.81-3.45 (m, 6H, CH₂NCH₂ and C(=O)CH₂CO₂), 3 98-4.15 (m, 4H, 2 x OCH₂CH₃), 5.05-5 12 (m, 1H, NCH=CHC(=O)), 6.69-7.50 m, 6H, C(2)H; C(4)H,

C(5)H, C(6)H, C(7)H indole and NCH=CH-C(=O)), 8.73 (br.s, 1H, NH indole). IR: 3480 (sh.s), 1730 (s), 1650 (m), 1600 (m), 1560 (s). MS exact mass: found 400.1982 (calculated for C₂₂H₂₈N₂O₅: 400.199).

Ethyl 5-[ethoxycarbonylethyl-(3-(N-methyl)indolyl)ethyl]amino-3-oxo-4-pentenoate (7 R1=Et).

Folate model 5 (750 mg, 2 mmol) and tryptamine derivative 3b (1410 mg, 5 mmol) were dissolved in a mixture of acetonitrile/acetic acid (16.5 ml, 10:1) and the mixture was refluxed for 30 h under a nitrogen atmosphere. The mixture was concentrated under vacuum and the residue was taken up in ethyl acetate and the solution was washed with sat. NaHCO3, brine and dried over MgSO4. After filtration silica was added and the solvent was carefully removed under reduced pressure. The remaining powder was brought on top of a silica gel column for flash chromatography (eluent ethyl acetate \rightarrow ethyl acetate/ethanol 1:1) and beside the product (433 mg, 52 %) also some starting material (5) was isolated (98 mg, 13 %). ¹H-NMR (200 MHz) broad signals due to hindered rotation: 0.91 and 0.97 (2 x t, 2 x 3H, J = 7.1 Hz, 2 x OCH₂CH₃), 1.85-2.10 (m, 2H, NCH₂CH₂CO₂), 2.60-3.40 (m, 11H, indole-CH₂-CH₂NCH₂, NCH₃ and C(=O)CH₂CO₂), 3.85 and 4.00 (2 x q, 2 x 2H, J = 7.1 Hz, 2 x OCH₂CH₃), 5.23 (br.d, 1H, NCH=CHC(=O)), 6.69-7.50 m, 6H, C(2)H, C(4)H, C(5)H, C(6)H, C(7)H indole and NCH=CHC(=O)). IR: 1730 (s), 1650 (m), 1600 (m), 1560 (s). MS exact mass: found 414.2140 (calculated for C₂₃H₃₀N₂O₅: 414.2154).

BF3 OE12 induced cyclization of enaminone 6a.

A mixture of 77 mg of 6a and 1 ml of freshly distilled BF₃·OEt₂ was stirred overnight at room temperature under a nitrogen atmosphere. After careful addition of sat. NaHCO₃, the water layer was extracted with dichloromethane. The combined organic layers were washed with brine, dried over MgSO₄ and concentrated under vacuum. The products were separated by chromatography on a silica plate (ethyl acetate) and 35 mg of the β -carboline 7a (45 %) and 15 mg of 8a (19 %) were isolated, both as a yellow oil. 2-Ethoxycarbonylethyl-1-(3-ethoxycarbonyl-2-oxo)propyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indole (7a R₁=Et).

¹H-NMR (200 MHz): 1.21-1.29 (m, 3H, CH₂CH₃), 2.49-2.59 (m, 4H, CH₂CH₂CO₂ and C(1)HCH₂C(=O)), 2.89-3.25 (m, 6H, 2 x C(3)H, 2 x C(4)H and CH₂C(=O)CH₂), 3.49 (s, 2H, C(=O)CH₂CO₂), 4.08-4.26 (m, 3H, C(1)H and OCH₂CH₃), 7.07-7.18 (m, 2H, C(6)H and C(7)H), 7.30 (d, 1H, J = 7.3 Hz, C(8)H), 7.47 (d, 1H, J = 8.2 Hz, C(5)H), 8.35 (br.s, 1H, NH). ¹³C-NMR (50 MHz, APT): 14.0 and 14.2 (2 x OCH₂CH₃), 17.9 (C(4)), 34.0 (CH₂CH₂CO₂), 45.2 (C(1)CH₂C(=O)CH₂), 49.0, 49.2 and 49.8 (C(3), C(=O)CH₂CO₂ and NCH₂CH₂CO₂), 52.6 (C(1)), 60.3 and 61.5 (2 x OCH₂CH₃), 107.7 (C(4a)), 110.9 (C(8)), 118.0 (C(5)), 119.2 (C(6)), 121.7 (C(7)), 126.8 (C(4b)), 134.0 and 135.7 (C(8a) and C(9a)), 116.8 (C(=O)CH₂CO₂), 172.4 (CH₂CH₂CO₂Et), 203.9 (CH₂C(=O)CH₂CO₂). **IR**: 3469 (sh.s), 1725 (s). MS exact mass: found 400.1993 (calculated for C₂2H₂8N₂O5 400.1998). *rel*-(3aS,6aR,11bS)-2.3.3a,4.6a,7-Hexahydro-5-hydroxy-6-methoxycarbonyl-3-(methoxy-carbonylethyl)-1H-pyrrolo[2,3-d]carbazole (8a R₁=Et).

¹H-NMR (200 MHz): 1.26 and 1.34 (2 x t, 2 x 3H, J = 7.1 Hz, 2 x OCH₂CH₃), 1.93 (dt, 1H, J = 9.1 and 13.5 Hz, C(6)Hg), 2.08-2.63 (m, 8H, C(3)H, C(5)H_Q, C(6)H_Q, 2 x C(14)H, 2 x C(20)H, C(21)H), 3.06 (dt, 1H, J = 8.1 and 11.8 Hz, C(3)H), 3.38-3.46 (m, 1H, C(5)Hg), 3.92-4.37 (m, 6H, 2 x OCH₂CH₃, C(16)H and C(2)H), 4.78 (d, 1H, J = 3.3 Hz, NH), 6.62 (d, 1H, J = 7.7 Hz, C(12)H), 6.78 (t, 1H, J = 7.4 Hz, C(11)H), 7.04-7.13 (m, 2H, C(9) and C(10)H). ¹³C-NMR (50 MHz, APT): 14.16 and 14.19 (2 x OCH₂CH₃), 34.0 (C(6)), 37.2 and 37.6 (C(20) and C(14)), 48.2 (C(3)), 52.9 (C(7)), 53.2 (C(5)), 54.4 (C(16)), 60.6 and 61.3 (OCH₂CH₃), 67.5 and 71.2 (C(2) and C(21)), 109.5 (C(12)), 119.2 (C(10)), 123.0 (C(9)), 128.5 (C(11)), 133.2 (C(8)), 150.0 (C(13)), 170.4 and 172.0 (2 x CO₂), 202.9 (C(17)). ¹H-NMR (C₆D₆, 250 MHz): 1.02 and 1.13 (2 x t, 2 x 3H, J = 7.1 Hz, 2 x OCH₂CH₃), 1.27 (dt, 1H, J = 9.1 and 13.7 Hz, C(6)Hg), 1.61-1.70 (m, 1H, C(14)H), 1.76 (dd, 1H, J = 9.1 and 12.2 Hz, C(6)H_Q), 1.93-2.32 (m, 6H, C(3)H, C(5)H_Q, C(14)H, 2 x C(20)H, C(21)H), 2.70 (ddd, 1H, J = 6.6, 10.0 and 12.0 Hz, C(3)H), 2.95-3.02 (m, 1H, C(5)H_B), 3.70-3.72 (dd, 1H, J = 1.8 and 3.3 Hz, C(2)H), 3.87-4.21 (m, 4H, 2 x OCH₂CH₃), 4.36 (d, 1H, J = 1.8 Hz, C(16)H), 4.79 (br.d, 1H, J = 3.3 Hz, NH), 6.25 (d, 1H, J = 7.0 Hz, C(12)H), 6.67-7.03 (m, 3H, C(9)H, C(10)H and C(11)H). IR: 3400 (sh.m), 2860 (m), 2810 (m), 1725 (s), 1710 (s), 1600 (m). MS exact mass: found 400.1997 (calculated for C₂₂H₂₈N₂O5: 400.1998).

2-Ethoxycarbonylethyl-1-(3-ethoxycarbonyl-2-oxo)propyl-9-methyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]undole (7b R1=Et).

Compound 6b (96 mg, 0.23 mmol) was dissolved in 2 ml of freshly distilled BF3·OEt2 and the mixture was heated to 60 °C. After refluxing for 5 h the reaction mixture was cooled on an ice-bath and sat. NaHCO3 was added carefully. The water layer was extracted with ethyl acetate and the combined organic layers were washed with brine, dried over MgSO4 and concentrated under vacuum. The product was obtained by chromatography on a silica plate and 47 mg of the B-carboline 7b was isolated as a yellow oil (49 %). ¹H-NMR (250 MHz): 1.25 and 1.28 (2 x t, 2 x 3H, J = 7.1 Hz, 2 x OCH₂CH₃), 2.50-2.58 (m, 4H, CH₂CH₂CO₂ and CHCH₂C(=O)CH₂), 2.65-3.19 (m, 6H, 2 x C(3)H, 2 x C(4)H, NCH₂CH₂CO₂, COCH₂CO₂), 3.60 (s, 3H, NCH₃), 4.06-4.23 (m, 4H, 2 x OCH₂CH₃), 4.32-4.37 (br.dd, 1H, C(1)H), 7.06-7.28 (m, 3H, C(6)H, C(7)H and C(8)H), 7.48 (d, 1H, J = 7.5 Hz, C(5)H).

R. H. HUIZENGA and U. K. PANDIT

¹³C-NMR (50 MHz, APT): 14.1 and 14.2 (2 x OCH₂CH₃), 16.8 (C(4)), 29.5 NCH₃), 33.8 (NCH₂CH₂CO₂), 43.0 (C(1)H₂CH₂-C(=O)CH₂), 47.0, 48.6 and 50.3 (C(3), C(=O)CH₂CO₂Et and NCH₂CH₂CO₂Et), 53.1 (C(1)), 60.3 and 61.3 (2 x O₂H₂CH₃), 107.3 (C(4a)); 108.8 (C(8)), 118.1 (C(5)), 119.1 (C(6)), 121.5 (C(7)), 126.7 (C(4b)), 134.2 and 137.1 (C(8a) and C(9a)), 167.2 and 172.4 (2 x O₂Et), 201.1 (CH₂C(=O)CH₂). IR: 1730 (br.s). MS exact mass: found 414.2160 (calculated for C_{23H30}N₂O₅ : 414.2155).

rel-(3a\$,6aR,11bR)-2,3,3a,4,6a,7-Hexahydro-5-hydroxy-6-methoxycarbonyl-3-(methoxy-carbonylethyl)-7-methyl-1H-pyrrolo-[2,3-d]carbazole (8b R1=Me).

Enaminone **6b** (165 mg, 0.43 mmol) was dissolved in 10 ml of dichloroethane and after addition of 1 ml of a 1.0 M TiCl4 solution in dichloroethane the mixture was refluxed overnight. The mixture was cooled to room temperature and a sat. NaHCO3 solution was added. The water layer was extracted with ethyl acetate and the combined organic layers were washed with brine and dried over MgSO4. After flash chromatography 84 mg of **8b** was isolated as a yellow oil (51 %). ¹H-NMR (200 MHz): 1.72-1.78 (m, 1H, C(6)Hg), 2.10-2.60 (m, 11H, C(3)H, C(5)H_{α}, C(6)H_{α}, 2 x C(14)H, 2 x C(20)H, C(21)H, NCH3), 3.02-3.23 (m, 2H, C(5)Hg and C(3)H), 3.67 (s, 3H, C(15)O₂CH₃), 3.80 (s, 3H, C(16)CO₂CH₃), 4.07 (s, 1H, C(2)H), 6.48 (d, 1H, J = 7.8 Hz, C(12)H), 6.73 (t, 1H, J = 7.3 Hz, C(11)H), 7.04-7.14 (m, 2H, C(9) and C(10)H), 12.66 (s, 1H, OH). ¹³C-NMR (50 MHz, APT): 31.5 (C(14)), 33.5 (C(6)), 33.7 (NCH3), 38.7 (C(20)), 48.8 (C(3)), 51.3 and 51.5 (2 x OCH₃), 53.1 and 53.9 (C(5) and C(7)), 69.6 and 73.5 (C(2) and C(21)), 93.9 (C(16)), 107.3 (C(12)), 118.6 (C(10)), 122.6 (C(9)), 127.8 (C(11)), 136.8 (C(8)), 152.2 (C(13)), 172.5 and 172.6 (z x CO₂), 179.1 (C(17)). **IR**: 2860 (m), 2810 (m), 1730 (s), 1645 (s), 1605(s). **MS** exact mass: found 386.1851 (calculated for C₂₁H₂₆N₂O₅ : 386.1841).

Ethyl 5-[(1,1-dimethyl-2-tosylamino)ethylmethyl]amino-2-ethyl-3-oxo-4-pentenoate (17).

The β -keto ester corresponding to 16 (1.58 g, 10 mmol) was added to a solution of 20 mmol of LDA in 80 ml of THF at 0 °C under a nitrogen atmosphere. The mixture was stirred for 30 min. and cooled to -78 °C and the imidazolidinium salt (3.94 g, 10 mmol) was added. The mixture was slowly warmed to 0 °C and after addition of 5 ml of sat. NH4Cl solution the solvent was removed under vacuum. The residue was taken up in ethyl acetate and the organic layer was washed with sat. NH4CO₃, washed with brine, dried over MgSO₄ and concentrated under vacuum. The residue was purified by flash chromatography (eluent ethyl acetate/petroleum ether 60-80 2:1) and the product was isolated as a white foam (1.663 mg, 39 %). ¹H-NMR (200 MHz): 0.93 (t, 3H, J = 7.1 Hz, CHCH₂CH₃), 1.25 (t, 3H, J = 7.1 Hz, OCH₂CH₃), 1.31 (s, 6H, NC(CH₃)₂CH₂), 1.82-1.92 (m, 2H, CHCH₂CH₃), 2.43 (s, 3H, ArCH₃), 2.71 (s, 3H, NCH₃), 2.95 (d, 2H, J = 6.8 Hz, HNCH₂C(CH₃)₂N), 3.27 (t, 1H, J = 7.5 Hz, COCHEtCO₂Et), 4.16 (q, 2H, J = 7.1 Hz, OCH₂CH₃), 5.15 (d, 1H, J = 12.3 Hz, NCH=CHCO), 5.64 (t, 1H, J = 6.8 Hz, CH₂NHTs), 7.30 (d, 2H, J = 8.2 Hz, C(3)H and C(5')H tosyl), 7.73 (d, 2H, J = 8.2 Hz, C(2)H and C(6')H tosyl), 7.86 (d, 1H, J = 12.3 Hz, NCH=CHCO). IR: 1725 (s), 1640 (m), 1600 (m), 1550 (s), 1340 (m), 1160 (s), 1090 (m). MS (FI 90 °C): 424.

4-Ethoxycarbonyl-1-[methoxycarbonylethyl-(2(3-(N-methyl)indolyl)ethyl)]amino-3-oxo-1-hexene (18).

The folate model 17 (942 mg, 2.22 mmol) and tryptamine derivative 3b (3.56 g) were dissolved in a mixture of 120 ml of acetonitrile/acetic acid (9:1) and the mixture was refluxed for 20 h under a nitrogen atmosphere. The solvent was removed under reduced pressure and the residue was taken up in ethyl acetate. The solution was washed with sat. NaHCO3 solution to remove the acid and with sat. NaCl solution. The organic layer was dried over MgSO4 and after concentration under vacuum the residue was purified by flash chromatography. 719 mg of the product was isolated as a yellow oil (75 % yield) and 2.600 g of the tryptamine derivative was recycled as its acetic acid salt (73 %). ¹H-NMR (200 MHz): 0.93 (t, 3H, J = 7.3 Hz, CHCH₂CH₃), 1.25 (t, 3H, J = 7,1 Hz, OCH₂CH₃), 1.79-1.96 (m, 2H, CHCH₂CH₃), 2.49-2.53 (br.m, 2H, NCH₂CH₂OC₂), 2.93-3.01 (m, 2H, indole-CH₂CH₂N), 3.25 (t, 1H, J = 7.3 Hz, CHCH₂CH₃), 3.40-3.52 (m, 4H, CH₂CH₂NCH₂CH₂), 3.67 and 3.74 (2 x s, 2 x 3H, NCH₃ and OCH₃), 4.16 (q, 2H, J = 7.1 Hz, OCH₂CH₃), 5.21 (m, 1H, NCCH=CHCO), 6.85 (s, 1H, C(2)H indole), 7.10-7.30 (m, 3H, C(5)H, C(6)H and C(7)H indole), 7.50-7.60 (m, 2H, C(4)H indole and NCH=CHCO). IR: 1730 (s), 1650 (m), 1600 (m), 1560 (s).

1-(3-Ethoxycarbonyl-2-oxo)pentyl-2-methoxycarbonylethyl-9-methyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indole (19).

To a solution of enaminone 18 (580 mg, 1.36 mmol) in 50 ml of dichloroethane, 2.7 ml of a 1.0 N TiCl₄ solution in CCl₄ was added and the solution was refluxed overnight. The mixture was cooled to room temperature and poured out in an ice cold sat. NaHCO3 solution. The organic layer was washed with brine, dried over MgSO4 and concentrated under vacuum. Flash chromatography (eluent ethyl acetate/petroleum ether 60-80 1:9 \rightarrow 9:1) furnished the product in 30 % yield as a yellow oil (174 mg). ¹H-NMR (200 MHz): 0.96 (t, 3H, J = 7.4 Hz, CHCH₂CH₃), 1.25 (t, 3H, J = 7.1 Hz, OCH₂CH₃), 1.83-1.98 (m, 2H, CHCH₂CH₃), 2.49-2.60 (m, 4H, CH₂CH₂N(R)CH₂CH₂CO₂Me and CHCH₂C(=O)CH), 2.62-3.20 (m, 6H, CH₂CH₂N(R)-

CH₂CO₂Me), 3.38-3.46 (m, 1H, CH₂CH₃), 3.59 and 3.66 (2 x s, 2 x 3H, NCH₃ and OCH₃), 4.19 (q, 2H, J = 7.1 Hz, OCH₂CH₃), 4.40 (m, 1H, CH₂CH_N), 7.05-7.51 (m, 4H, C(4)H, C(5)H, C(6)H and C(7)H indole). **IR**: 1730 (s), 1705 (s), 1175 (m). MS exact mass: found 428.3213 (calculated for C₂₄H₃₂N₂O₅: 428.3231).

Methyl (1S)-3-(3-indolyl)-2-(2-methoxycarbonylethyl)aminopropanoate (28).

A solution of 3.18 g of the HCl-salt of the methyl ester of L-(-)-tryptophane 27 in chloroform was washed with a K₂CO₃ solution. The solution was dried over MgSO₄ and after concentration under vacuum the residue was dissolved in methanol. Methyl acrylate (1 eq) was added dropwise to the solution, which was stirred during 30 h at room temperature under a nitrogen atmosphere. After concentration under vacuum and flash chromatography (ethyl acetate/methanol) 2.35 g of the product was isolated as a yellow oil (62 %). ¹H-NMR (250 MHz): 1.98 (br.s, 1H, NH), 2.45 (t, 2H, J = 6.6 Hz, NCH₂CH₂CO₂Me), 2.70-2.99 (m, 2H, NCH₂CH₂CO₂Me), 3.03-3.24 (m, 2H, indole-CH₂), 3.57 and 3.64 (2 x s, 2 x 3H, 2 x CO₂CH₃), 3.57-3.68 (m, 1H, indole-CH₂CH₂CH₂CH₂CO₁M), 7.59 (d, 1H, J = 2.0 Hz, C(2)H indole), 7.07-7.20 (m, 2H, C(5)H and C(6)H indole), 7.31 (d, 1H, J = 7.4 Hz, C(7)H indole), 7.59 (d, 1H, J = 7.3 Hz, C(4)H indole), 8.34 (br.s, 1H, NH indole). IR: 3480 (s), 3320 (br.w), 1730 (s).

Ethyl 5[(1,1-dimethyl-2-tosylamino)ethylmethyl]amino-3-oxo-4-pentenoate (29).

To a solution of diisopropylamine (3.050 g, 30.2 mmol) in 80 ml of dry THF at 0 $^{\circ}$ C under a nitrogen atmosphere was added 20 ml of 1.51 N BuLi solution in hexane. After addition of 15.6 mmol of ethyl acetoacetate the orange solution was cooled to -78 $^{\circ}$ C and the imidazolidinium salt 15 (6.107 g, 15.5 mmol) was added. The reaction mixture was allowed to warm up to 0 $^{\circ}$ C in 5 h upon which the mixture became clear. The reaction was quenched by addition of 2 ml sat. NH4Cl solution and after stirring for 15 min the solvent was removed under reduced pressure, the residue was taken up in ethyl acetate, the layers were separated and the organic layer was washed with brine, dried over MgSO4 and filtered. Silica gel was added and the solvent was removed carefully under vacuum. The powder was brought on top of a silica gel column for flash chromatography (eluent ethyl acetate/petroleum ether 60-80 1:1 \rightarrow ethyl acetate). Only the ring-opened form of the product was isolated as a yellow foam in 90 % yield (5.43 g). ¹H-NMR (250 MHz): 1.18 (t, 3H, J = 7.1 Hz, OCH₂CH₃), 1.31 (s, 6H, NC(CH₃)₂CH₂), 2.30 (s, 3H, ArCH₃), 2.82 (s, 3H, NCH₃), 3.44 (s, 2H, COCH₂EtCO₂Me), 3.50-3.70 (m, 2H, CH₂NH), 4.03 (q, 2H, J = 7.1 Hz, OCH₂CH₃), 5.16 (d, 1H, J = 11.9 Hz, NCH=CHCO), 6.78 (br.s, 1H, CH₂NHTs), 7.17 (d, 2H, J = 8.1 Hz, C(3)H and C(5)H tosyl), 7.75 (d, 2H, J = 8.1 Hz, C(2)H and C(6)H tosyl), 7.98 (d, 1H, J = 11.9 Hz, NCH=CHCO). IR: 1735 (s), 1635 (m), 1595 (m), 1550 (s), 1160 (s), 1090 (m).

Methyl (1S)-3-(3-indolyl)-2-([1-(4-ethoxycarbonyl-3-oxo-1-butene)](2-methoxycarbonylethyl))aminopropanoate (30).

Folate model 29 (1.4 mmol, 548 mg) and 1.1 eq tryptophane derivative 28 (480 mg) were refluxed overnight in a mixture of 10 ml of acetonitrile and 1 ml of acetic acid under a nitrogen atmosphere. After concentration under vacuum a solution of the residue in CH₂Cl₂ was washed with sat. NaHCO₃ solution and brine. The organic layer was dried over MgSO₄ and concentrated under vacuum. After flash chromatography (ethanol/dichloromethane 1:19) 211 mg of the product was isolated (34 %) as a brown oil. 160 mg of the starting tryptophane derivative 28 (33 %) could be recycled. ¹H-NMR (250 MHz): 1.27 (t, 3H, J = 7.2 Hz, OCH₂CH₃), 2.38-2.50 (m, 2H, NCH₂CH₂CO₂Me), 3.17-3.53 (m, 6H, NCH₂CH₂CO₂Me, indole-CH₂, COCH₂CO₂Et), 3.58 and 3.74 (2 x s, 2 x 3H, 2 x CO₂CH₃), 4.18 (q, 2H, J = 7.2 Hz, OCH₂CH₃), 4.35 (dd, 1H, J = 5.4 and 10.0 Hz, indole-CH₂CH₃), 5.16 (d, 1H, J = 13.1 Hz, NCH=CH₂CO), 6.97 (d, 1H, J = 2.3 Hz, C(2)H indole), 7.10-7.24 (m, 2H, C(5)H and C(6)H indole), 7.36 (d, 1H, J = 7.1 Hz, C(7)H indole), 7.56-7.64 (m, 2H, C(4)H indole and NCH=CHCO), 8.28 (br.s, 1H, NH indole). IR: 3480 (s), 3320 (br.w), 1730 (s), 1650 (m), 1600 (m), 1560 (s). MS (FD 10 mA): 444. [α]²⁰D = -121^o (CHCl₃, c = 0.0102 g/ml).

(2S,3aR,6aS,11bS)-6-Ethoxycarbonyl-2,3,3a,4,6a,7-hexahydro-5-hydroxy-2-methoxycarbonyl-3(2-methoxycarbonyl)ethyl-1H-pyrrolo[2,3-d]carbazole (31).

Compound 30 (146 mg) was dissolved in 15 ml of dichloroethane and 600 μ l of a 1M solution of TiCl4 in CCl₄ was added dropwise. After refluxing for 5 h and cooling to room temperature, the reaction mixture was added to a sat. NaHCO₃ solution. The mixture was extracted with dichloromethane. The organic layer was washed with brine and dried over MgSO₄. After concentration under vacuum the residue was chromatographed (ethyl acetate/petroleum ether 60-80 1:2) 69 mg of the product was isolated as a colourless oil (46 %). ¹H-NMR (250 MHz): 1.34 (t, 3H, J = 7.1 Hz, OCH₂CH₃), 2.08 (dd, 1H, J = 5.8 and 14.4 Hz, C(6)H_β), 2.34 (dd, 1H, J = 3.0 and 18.8 Hz, C(20)H), 2.40 (br.s, 2H, NCH₂CH₂CO₂Me), 2.54 (dd, 1H, J = 2.8 and 18.8 Hz, C(20)H), 2.40 (br.s, 2H, NCH₂CH₂CO₂Me), 2.54 (dd, 1H, J = 2.8 and 18.8 Hz, C(20)H), 2.85 (m, 1H, C(21)H), 2.90 (dd, 1H, J = 10.5 and 14.4 Hz, C(6)H_Q), 3.08 (dt, 1H, J = 7.8 and 13.5 Hz, C(3)H), 3.53 (dd, 1H, J = 5.8 and 10.5 Hz, C(5)H), 3.53 and 3.64 (2 x s, 2 x 3H, 2 x CO₂CH₃), 4.15-4.20 (m, 1H, C(2)H), 4.21-4.35 (m, 2H, OCH₂CH₃), 4.90-4.93 (m, 2H, NH and C(16)H), 6.59 (d, 1H, J = 7.7 Hz, C(12)H), 6.73 (t, 1H, J = 7.3 md)

Hz, C(10)H), 7.04-7.09 (m, 2H, C(9)H and C(12)H). IR: 3320 (br.w), 1730 (br.s), 1600 (m). MS (FD 10 mA): 444. $[\alpha]^{20}D = -50.3^{\circ}$ (CHCl₃, c = 0.0167 g/ml).

References and notes

- 1 for part 21 see: Huizenga, R.H.; van Wiltenburg, J.: Pandit, U.K., Tetrahedron Lett. 1989, 30, 7105.
- 2 Taken in part from the doctorate dissertation of R.H. Huizenga, University of Amsterdam, 1990.
- a) See: Pandit, U.K. Recl.Trav.Chim.Pays-Bas, 1988, 107, 111 and references cited therein;
 b) Stoit, A.R.; Pandit, U.K. Tetrahedron 1989, 45, 849.
- 4 Pandit, U.K.; Bieräugel, H.; Stoit, A.R. Tetrahedron Lett. 1984, 25, 1513.
- 5 For a review see: Saxton, E.J. in "The Chemistry of Heterocyclic Compounds", Vol. 25, pt. 4, "Indoles, The Monoterpenoid Indole Alkaloids", p.331-438, Saxton, J.E. (ed.), Wiley, New York (1983) and references cited therein.
- a) Ando, M.; Büchi, G.; Ohnuma, T. J.Am.Chem.Soc. 1975, 97, 6880; b) Wenkert, E.; Orito, K.;
 Simmons, D.P.; Kunesch, N.; Ardisson, J.; Poisson, J. Tetrahedron 1983, 39, 3719; c) Wenkert, E.;
 Porter, B.; Simmons, D.P.; Ardisson, J.; Kunesch, N.; Poisson, J. J.Org.Chem. 1984, 49, 3733; d)
 Blowers, J.W.; Brennan, J.P.; Saxton, J.E. J.C.S. P. I 1987, 2079; e) Jackson, A.H.; Shannon,
 P.V.R.; Wilkins, D.J. Tetrahedron Lett. 1987, 28, 4901.
- 7 Numbering as in aspidospermidine:



- 8 The tetracyclic products of the cyclization reactions were isolated as mixtures of keto-enol tautomers. The spectroscopic data of the predominant tautomer (more than 90%) are given in the experimental section.
- 9 Bailey, P.D. J.Chem.Res.(S) 1987, 202.
- 10 Still, W.C.; Kahn, M.; Mitra, A. J. Org. Chem. 1987, 43, 2923.